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RESEARCH ARTICLE

Ultraviolet and yellow reflectance but not fluorescence is important for visual discrimination of conspecifics by *Heliconius erato*

Susan D. Finkbeiner^{1,2,3,*}, Dmitry A. Fishman⁴, Daniel Osorio⁵ and Adriana D. Briscoe^{1,*}

ABSTRACT

Toxic *Heliconius* butterflies have yellow hindwing bars that – unlike those of their closest relatives – reflect ultraviolet (UV) and long wavelength light, and also fluoresce. The pigment in the yellow scales is 3-hydroxy-DL-kynurenine (3-OHK), which is found in the hair and scales of a variety of animals. In other butterflies like pierids with color schemes characterized by independent sources of variation in UV and human-visible yellow/orange, behavioral experiments have generally implicated the UV component as most relevant to mate choice. This has not been addressed in *Heliconius* butterflies, where variation exists in analogous color components, but moreover where fluorescence due to 3-OHK could also contribute to yellow wing coloration. In addition, the potential cost due to predator visibility is largely unknown for the analogous well-studied pierid butterfly species. In field studies with butterfly paper models, we show that both UV and 3-OHK yellow act as signals for *H. erato* when compared with models lacking UV or resembling ancestral *Eueides* yellow, respectively, but attack rates by birds do not differ significantly between the models. Furthermore, measurement of the quantum yield and reflectance spectra of 3-OHK indicates that fluorescence does not contribute to the visual signal under broad-spectrum illumination. Our results suggest that the use of 3-OHK pigmentation instead of ancestral yellow was driven by sexual selection rather than predation.

KEY WORDS: Visual signal, Yellow pigment, Sexual selection, Predation, Mate preference, Light environment

INTRODUCTION

Color patches of animals are complex traits composed of multiple components (Grether et al., 2004). The pigment cells known as chromatophores in the skin of fishes, reptiles and amphibians, for example, are color-generating structures composed of distinct pigmentary and structural layers that vary in their ability to reflect light. The feather barbs or integument of birds or the wing scales of butterflies similarly have diverse nano-structure architectures, thin films and pigments, which produce a dazzling array of colors (Prum and Torres, 2003; Vukusic and Sambles, 2003; Shawkey and Hill, 2005; Stavenga et al., 2011, 2014). These pigmentary and structural components of color patches work in tandem to produce signals used in a variety of contexts (e.g. crypsis, mimicry, aposematism and mate

choice). Because the biochemical and developmental mechanisms underlying the pigmentary and structural properties of color differ, each of these components may be subject to different selective pressures and hence independent evolutionary trajectories (Grether et al., 2004). Here, we looked specifically at how two components of a butterfly visual display – UV reflectance and human-visible yellow reflectance due to selective filtering by a specific wing pigment – may function as a signal in mate choice and predation. We also looked at what contribution fluorescence makes, if any, to the signal.

Many butterfly species have colorful wing patterns in both the human-visible (400–700 nm) and UV (300–400 nm) ranges (Silberglied and Taylor, 1978; Eguchi and Meyer-Rochow, 1983; Meyer-Rochow, 1991; Rutowski et al., 2005; Briscoe et al., 2010). While the idea that UV coloration – invisible to humans – may serve as a ‘private channel’ of communication has been challenged (Cronin and Bok, 2016; but see Cummings et al., 2003), there is ample evidence that UV signals are important in animal communication (Rutowski, 1977; Johnsen et al., 1998; Smith et al., 2002; Cummings et al., 2003; Robertson and Monteiro, 2005; Kemp, 2008; Obara et al., 2008; Detto and Blackwell, 2009; Painting et al., 2016). However, although many butterflies have UV-visible color patches, in the absence of behavioral evidence, it is unclear whether the UV reflectance functions as a signal or whether it is simply an epi-phenomenon of the scale structure overlaying pigment granules. The same question can of course be applied to the colors produced by the pigments.

Studies of several butterfly groups suggest in fact that for color patches with both UV and visible reflectance, only variation in the UV component of the signal affects mate choice. Pierid butterfly males, *Colias eurytheme* and *Colias philodice*, have forewing colors with both UV iridescence due to the structural scattering of light by the scale lamellae (Ghiradella, 1974) and yellow–orange reflectance due to pterin pigments (Watt, 1964). In behavioral experiments, female *Colias* were shown to use the UV-reflection difference between the two species as a mate and species recognition cue, but not the human-visible color difference (Silberglied and Taylor, 1978). Female *Eurema hecuba* (Coliadinae: Pieridae) were similarly shown to prefer males with the brightest UV iridescence overlaying a diffuse pigment-based yellow (Kemp, 2007a). Given that many other butterflies have color patches with UV reflectance, and that butterfly color vision systems are astonishingly diverse (Arikawa et al., 2005; Briscoe and Bernard, 2005; Stalleicken et al., 2006; Koshitaka et al., 2008; Sison-Mangus et al., 2008; Chen et al., 2013), it is worthwhile investigating in other species whether it is the UV or the human-visible part of the color patch reflectance spectrum, or both, that is being used for signaling. It is particularly interesting to investigate this question where there has been a phylogenetic transition from using one type of pigmentation to another, as for the yellow wing colors in the passion-vine butterflies of the genus *Heliconius* (Briscoe et al., 2010; Bybee et al., 2012) (see below).

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Heliconius erato has yellow scales on its hindwings that contain the pigment 3-hydroxy-DL-kynurenine (3-OHK) (Tokuyama et al., 1967; Reed et al., 2008). The yellow bars reflect UV light and have a step-like reflectance at longer wavelengths – a rapid rise then a plateau in reflectance in the visible (400–700 nm) range (Fig. 1A,B, yellow lines) (see also Stavenga et al., 2004). Either the UV or the human-visible part of 3-OHK wing reflectance, or both, may serve as a signal for inter- and intra-specific communication. Intriguingly, the appearance of 3-OHK in *Heliconius* co-occurred with the evolution of the butterflies' duplicated UV opsins, UV1 and UV2 (Briscoe et al., 2010; Yuan et al., 2010; Bybee et al., 2012). In some *Heliconius* species, UV1 and UV2 are found in both males and females (K. J. McCulloch and A.D.B., unpublished data). In *H. erato*, UV1 is a female-specific UV receptor with $\lambda_{\text{max}}=355$ nm, while UV2 is a violet receptor found in both sexes with $\lambda_{\text{max}}=390$ nm (McCulloch et al., 2016).

In addition to the components of the 3-OHK visual signal mentioned above, the yellow wing bars of *Heliconius* fluoresce under a hand-held blacklight (Movie 1). Fluorescence occurs when short-wavelength light is absorbed and then re-emitted as a longer wavelength, i.e. lower energy, light. Fluorescent pigments are widespread in nature (Vukusic and Hooper, 2005; Lagorio et al., 2015) and are typically identified using spectrally narrow-band light; however, terrestrial illumination has a broad spectrum so it is unclear whether a pigment's fluorescence contributes much to a

potential signal under natural conditions. The emission spectra of the 3-OHK pigment overlap with the visible part of the reflectance spectrum of 3-OHK on *Heliconius* wings (see below) and so would be well suited to being detected by the blue-sensitive receptor of *H. erato* with $\lambda=470$ nm if it did (McCulloch et al., 2016).

Butterflies from the genus *Eueides*, which is a sister taxon to *Heliconius*, have mimetic wing patterns strikingly similar to those of some *Heliconius* species. These two genera co-occur in the same habitats, yet *Eueides*' yellow wing pigments lack the step-like reflectance spectrum of 3-OHK (Fig. 1A,B, gray line) (Bybee et al., 2012), and they do not fluoresce (data not shown). The yellow pigments in the two butterflies appear similar to the human eye in natural light, but their spectra differ strongly (Fig. 1A,B, yellow and gray lines). Although modeling of wing colors suggests in principle that *Heliconius* can distinguish between *Heliconius* 3-OHK yellow and *Eueides* yellow (Bybee et al., 2012), it remains unknown whether *Heliconius* actually do so in nature. Previous work has shown that *H. erato* prefer chromatic over achromatic signals in the context of mate choice (Fig. S1) (Finkbeiner et al., 2014); but it is unclear whether the visible, the UV or both parts of the reflectance spectrum of 3-OHK and fluorescence contribute to signaling. Prior work has also shown that avian predators will differentially attack achromatic compared with chromatic butterfly paper models (Fig. S1) (Finkbeiner et al., 2014; Dell'Aglio et al., 2016), but it is unknown whether avian predators will differentially attack butterfly

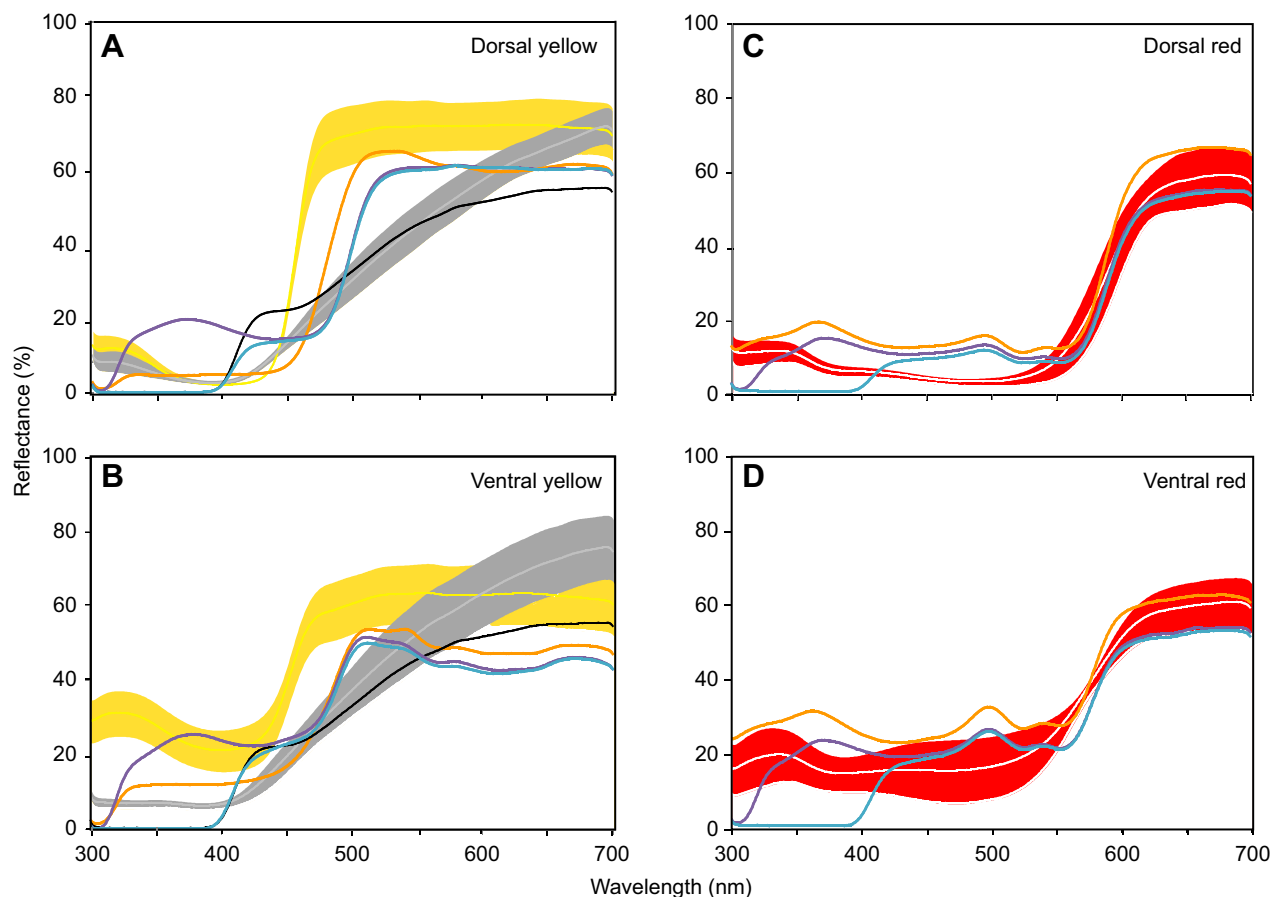


Fig. 1. Reflectance spectra of *Heliconius erato* and *Eueides* wing colors and paper model colors used in the mate choice and predation experiments. (A) Dorsal yellow, (B) ventral yellow, (C) dorsal red, (D) ventral red. Reflectance spectra correspond to: natural *H. erato* wing colors (yellow or red; $n=15$ butterflies), natural *Eueides* spp. wing colors (gray; $n=11$ butterflies) and paper model wing colors with (Y+, orange) and without (Y-, black) 3-OHK color, and with (UV+, purple) and without (UV-, blue) ultraviolet reflectance. Shaded areas correspond to 95% confidence intervals; solid yellow or white lines are means with (Y+) and without (Y-) 3-OHK yellow, and with (UV+) and without (UV-) ultraviolet reflectance.

paper models that vary in yellow coloration resembling the differences between *Heliconius* and *Eueides* yellow. While *Heliconius* wing color patterns warn avian predators of their toxicity (Benson, 1972; Chai, 1986), 3-OHK may further serve as a conspecific signal especially in courtship (Bybee et al., 2012; Llaurens et al., 2014). Demonstrating that *Heliconius* species can in fact discriminate 3-OHK yellow from other yellows in nature is an important step in elucidating the adaptive significance of 3-OHK pigmentation.

To further investigate the contribution of 3-OHK to *H. erato* signaling, we carried out two sets of experiments. The first set of experiments tested responses of both male and female *H. erato* to four types of colored models with spectra that were intended to approximate those of either *Heliconius* species or their mimics, such as *Eueides*. The first pair of spectra, which are designated Y+ or Y–, resemble 3-OHK (*Heliconius*) yellow or *Eueides* yellow, respectively; the second set of reflectance spectra have identical yellow and red coloration in the visible range, but UV reflectance is either present (UV+) or absent (UV–). The second, complementary set of experiments tested the hypothesis that predatory birds will not differentially attack 3-OHK yellow from other yellows when presented with model butterflies as a result of the aposematic function of yellow in general.

Together, these experiments substantiate and elaborate our understanding of the function of 3-OHK yellow and UV coloration. We show also that fluorescence – although clearly visible in laboratory conditions, but with illumination restricted to the UV excitation wavelengths – is not likely to have any impact under the broadband and relatively low UV illumination found in nature.

MATERIALS AND METHODS

Butterfly models, wing reflectance spectra, environmental light and discriminability

Four paper model types of the *Heliconius erato petiverana* Doubleday 1847 butterfly were made as described in Finkbeiner et al. (2012) with their colors modified as follows: with (Y+) and without (Y–) 3-OHK yellow, and with (UV+) and without (UV–) ultraviolet reflectance. The Y+ treatment had 3-OHK on the yellow portion of the wing (0.010 and 0.015 mg μl^{-1} 3-OHK in methanol applied to the ventral and dorsal sides, respectively). This provided the models with the same pigment as found in the butterfly yellow scales (Fig. 1A,B, orange lines). The yellow portion of the non-3-OHK yellow models (Y–) was covered with yellow Manila paper (Creatology® Manila Drawing Paper, item no. 410590). Manila paper has a reflectance spectrum that resembles non-3-OHK yellow reflectance from the sister genus to *Heliconius*, *Eueides*, which is a *Heliconius* mimic (Bybee et al., 2012) (Fig. 1A,B, gray and black lines). A thin film UV filter (Edmund Optics, item no. 39-426) was placed over the Manila paper to create a closer match to *Eueides* yellow pigment. As a control, Mylar film was added to the yellow portions of models with 3-OHK for the Y+ treatment. Mylar film resembles the UV filter but acts as a neutral-density filter. The red portions of the wings were identical in Y+ and Y– treatments.

For the UV+ models, an odorless UV-reflective yellow paint (Fish Vision™) was added to the dorsal and ventral yellow band of the model wings to provide UV reflectance (Fig. 1A,B, purple line), and the red portions of the wings were printed as described in Finkbeiner et al. (2014). For UV models, a thin film UV filter was placed over both the yellow and red/pink UV-reflective portions on the wings. The UV filter prevents any light reflectance up to 400 nm (Fig. 1, blue line). Mylar film was added to the yellow and red/pink

portions of models used for the UV+ treatment to function as a control.

Reflectance spectra of the paper models and individual *Heliconius erato petiverana* ($n=15$), *Eueides isabella*, *E. surdus*, *E. thales* ($n=3$ per species) and *E. heliconoides* ($n=2$) butterfly wings were measured by first aligning each measured wing in the same orientation as shown in appendix B of Bybee et al. (2012). If the viewer was looking directly from above at the oriented wings, the fixed probe holder (Ocean Optics RPH-1) was placed horizontally on top of the wing such that the axis of the illuminating and detecting bifurcating fiber (Ocean Optics R400-7-UV/VIS) was at an elevation of 45 deg to the plane of the wing and pointed left with respect to the body axis. Illumination was by a DH-2000 deuterium–halogen lamp, and reflectance spectra were measured with an Ocean Optics USB2000 spectrometer. A spectralon white standard (Ocean Optics WS-1) was used to calibrate the spectrometer. For the irradiance spectra measurements, the USB2000 spectrometer, a calibrated tungsten light source (Ocean Optics LS-1-CAL), a 100 or 400 μm diameter fiber (Ocean Optics P100- or P400-2-UV-Vis) and cosine corrector (Ocean Optics CC-3-UV), which produces vector irradiance measures, were used (Cronin et al., 2014). Five irradiance spectra measurements of down-dwelling light were taken and averaged per site.

For the mate-choice experiments, the von Kries-transformed quantum catches for stimuli (Kelber et al., 2003) were first calculated for *H. erato* males and females separately using high light intensity and sunny cage irradiance spectra. Pairwise discriminabilities between artificial models and natural wing reflectance spectra were determined using a trichromatic vision model for *H. erato* males and tetrachromatic vision models for *H. erato* females (Vorobyev and Osorio, 1998). Parameters for the butterfly visual models were as follows: Weber fraction=0.05 (Koshitaka et al., 2008); photoreceptor peak sensitivity, λ_{max} , of 355 nm (female only), 390 nm, 470 nm and 555 nm; and relative abundance of photoreceptors, violet-sensitive (VS)=0.13, B=0.2, G=1 (male) or UV=0.09, VS=0.07, B=0.17, G=1 (female) (McCulloch et al., 2016). For the predation experiments, von Kries-transformed quantum catches for only ventral wing stimuli (as the butterflies were presented with their wings folded) were calculated using high light intensity and irradiance spectra from two of the four habitats where the models were placed: forest cover and forest edge. (The other two habitats, Pipeline Road and paved road, were found to have normalized spectra that were identical to forest cover.) Discriminabilities between stimuli were determined using tetrachromatic models of bird vision representing two types of avian visual system, the UV-type (blue tit, *Cyanistes caeruleus*) and violet-type (chicken, *Gallus gallus*) systems (reviewed in Frentiu and Briscoe, 2008). For chicken, we used ocular media of Lind and Kelber (2009) and Toomey et al. (2016) and behaviorally determined parameters of Olsson et al. (2015); namely, a Weber fraction of 0.06 for the L cone, and relative abundance of cones: VS=0.25, S=0.5, M=1, L=1. For the blue tit, we followed the work of Hart et al. (2000) including the effects of blue tit ocular media and used a Weber fraction of 0.05 for the L cone, and relative abundance of cones: UV=0.37, S=0.7, M=0.99, L=1.

Mate preference experiments

To test whether *Heliconius* 3-OHK yellow and UV serve as visual signals for conspecifics, mate preference experiments were carried out using insectary facilities in Gamboa, Panama, from September 2013 to February 2014. Data were collected from 80 wild-caught *H. erato petiverana* butterflies: 40 males and 40 females. Each

butterfly was introduced individually into experimental cages (2 m×2 m×2 m) and presented with one of two pairs of the artificial butterfly models: Y+ versus Y−, or UV+ versus UV−. The models were separated by 1 m and attached to an apparatus used to simulate flight (see Finkbeiner et al., 2014). Movies 2 and 3 show an example of female butterfly trials with Y (Movie 2) and UV (Movie 3) models. Individual butterflies experienced six 5 min trials – three 5 min trials with each of the two pairs. During trials, two variables were recorded: (1) approaches, which consisted of flight unequivocally directed toward the model, and in which the butterfly came within 20 cm of the model, and (2) courtship events, which were classified as sustained hovering or circling behavior around the model (for examples, see videos 2 and 3 in Finkbeiner et al., 2014). Mate preference data were analyzed using a two-way ANOVA in R to examine the effects of model type and sex. Measurements of spectral irradiance (see above) were taken to provide quantitative information about the illumination conditions during the trials (Fig. S2).

Predation experiments

Previously, we have shown (Finkbeiner et al., 2014) that avian predators differentially attack achromatic local-form butterfly models compared with chromatic models as well as models that display non-local or color-switched patterns (Fig. S1). Here, we tested whether avian predators would differentially attack local wing color form paper models where UV or yellow is manipulated. Predation experiments were completed in Panama at the Smithsonian Tropical Research Institute Gamboa field station and at selected forest sites in Soberanía National Park (including Pipeline Road), from June to September in 2013. Models were fitted with Plasticine abdomens and tied to branches with thread to represent natural resting postures in the following habitat types: forest cover (15 sites), forest edge (17 sites), Pipeline Road (unpaved road with partial forest cover, 55 sites) and paved road with partial forest cover (13 sites). Examples of foliage cover in each of these habitat types, along with corresponding spectral irradiance measurements, are presented in Fig. S3. For the 3-OHK yellow pigment study, five artificial models of each treatment (Y+ and Y−) were randomly placed in 100 forest sites (Finkbeiner et al., 2014). The sites were separated by ~250 m to account for avian predator home range (home ranges described in Finkbeiner et al., 2012). There were 500 Y+ models and 500 Y− models for a total of 1000 models. The same methods were used for the UV study, using 500 UV+ models and 500 UV− models in non-overlapping sites from the Y± models, and with the same number of habitat types.

The models remained at their sites for 4 days, and each model was examined for evidence of predation. A butterfly was considered attacked if damage to the abdomen and wings appeared in the form of beak marks and/or large indentations in the abdomen (for examples of attacked models, see Finkbeiner et al., 2012, 2014). The attack response was modeled as a binomial variable (yes or no) dependent upon butterfly model type using a zero-inflated Poisson regression model, including sites as a random effect, in R with the ‘pscl’ package (Zeileis et al., 2008; R Development Core Team, 2010; Jackman, 2011). To examine whether forest light environment affected predator behavior, the same analysis was used to compare predation between model types in four main habitat types: forest cover, forest edge, Pipeline Road (unpaved road with partial forest cover) and paved road with partial forest cover.

Fluorescence experiments

To determine the possible contribution of 3-OHK fluorescence to its yellow coloration, we measured the absorption, excitation and emission spectra of 1.5 mg 3-hydroxy-DL-kynurenine (3-OHK; Sigma-Aldrich, catalog no. H1771) in 3 ml methanol (Fisher Chemicals, Optima LC/MS grade, catalog no. A456-1). The resultant solution was diluted to an optical density (OD)=0.3 to get it within the linear range for fluorescence measurement (Dhami et al., 1995). The absorption spectrum of the pigment was measured with a Cary-50 spectrometer (Varian), while the emission and excitation spectra were acquired with a Cary Eclipse fluorimeter (Varian).

We determined the quantum yield of 3-OHK pigment using the comparative method of Williams et al. (1983). The method makes use of a well-characterized standard with a known quantum yield and an absorbance spectrum that is similar to the absorbance spectrum of the sample of interest, in this case 3-OHK. When the reference and the sample of interest have a similar absorbance at the fluorescence excitation wavelength, the amount of photons being absorbed by the reference and test solutions can be assumed to be the same. In this case, a simple ratio of integrated fluorescence is equal to the ratio of the quantum yields of the reference and sample of interest. For greater accuracy, six additional experiments were performed using solutions of various absorbances (ODs). The integrated fluorescence intensity was then plotted against the absorbance of each solution and if this represented a linear function, where no reabsorption occurred, then the measurement was retained; otherwise, the experiment was discarded. The ratio of the slopes of these functions for the reference and sample of interest

Table 1. Percentage of *Heliconius erato* and *Eueides* wing colors compared with paper models with chromatic JND values <0.5, <1 or <2 for male and female *H. erato* under high light, sunny cage illumination

		Y+		Y−		UV+		UV−	
		Female	Male	Female	Male	Female	Male	Female	Male
Dorsal yellow	0.5 JND	0.0	6.7	0.0	0.0	0.0	0.0	36.4	63.6
	1 JND	86.7	100.0	81.8	9.1	0.0	0.0	100.0	100.0
	2 JND	100.0	100.0	100.0	100.0	0.0	0.0	100.0	100.0
Ventral yellow	0.5 JND	13.3	0.0	0.0	55.6	33.3	33.3	0.0	11.1
	1 JND	86.7	86.7	100.0	77.8	100.0	100.0	88.9	66.7
	2 JND	100.0	100.0	100.0	100.0	100.0	100.0	100.0	100.0
Dorsal red	0.5 JND	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
	1 JND	13.3	0.0	13.3	0.0	6.7	0.0	0.0	0.0
	2 JND	86.7	46.7	86.7	46.7	86.7	46.7	93.3	80.0
Ventral red	0.5 JND	46.7	46.7	46.7	46.7	46.7	46.7	0.0	0.0
	1 JND	46.7	46.7	46.7	46.7	46.7	46.7	0.0	0.0
	2 JND	100.0	66.7	100.0	66.7	100.0	73.3	60.0	100.0

Values are percentage below threshold. JND, just-noticeable difference. *n*=15 *H. erato*, *n*=9 *Eueides* specimens measured.

Table 2. Percentage of *H. erato* and *Eueides* wing colors compared with paper models with chromatic JND values <0.5, <1 or <2 for the UV-type blue tit (*Cyanistes caeruleus*) and violet-type chicken (*Gallus gallus*) visual systems under high light, partial forest cover (or forest edge) illumination

		Y+		Y–		UV+		UV–	
		UV	VS	UV	VS	UV	VS	UV	VS
Ventral yellow	0.5 JND	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
	1 JND	0.0	0.0	0.0	66.7	33.3	0.0	0.0	0.0
	2 JND	86.7	6.7	88.9	77.8	100.0	100.0	0.0	0.0
Ventral red	0.5 JND	0.0	20.0	0.0	20.0	6.7	13.3 (20.0)	0.0	0.0
	1 JND	33.3	46.7	33.3	46.7	33.3	46.7	0.0	13.3
	2 JND	46.7	46.7	46.7	46.7	46.7	46.7	0.0	46.7

VS, violet sensitive. Values are percentage below threshold and were identical for partial forest cover and forest edge illumination except for the number indicated in parentheses. $n=15$ *H. erato*, $n=9$ *Eueides* specimens measured.

is equal to the quantum yield ratio. For this particular experiment, Coumarin 500 (Exciton, catalog no. 05000) was chosen as a reference as its emission and absorption spectrum are extremely similar to those of 3-OHK.

The reflectance spectrum measurements of *H. erato* wings were made using an Ocean Optics USB2000 spectrometer, a UV-cut off filter (Edmund Optics no. 39-426), a 150 W Xenon Arc lamp (which resembles daylight illumination) and a Spectralon white standard.

RESULTS

Discriminabilities of model spectra and real wings

To test the hypothesis that our Y+ and UV+ paper models resembled real *H. erato* yellow wing colors, and that our Y– and UV– paper models resembled real *Eueides* yellow wing colors, we calculated pairwise discriminabilities between real wings and model spectra. We did this for the male and female *H. erato* visual system, and then for the UV-type and VS-type avian visual systems. We found that for both male and female *H. erato* eyes, Y+ was an excellent match to *H. erato* dorsal and ventral yellows, and that Y– and UV– were excellent matches to *Eueides* dorsal and ventral yellows under high light illumination [Table 1; 66.7–100% of pairwise comparisons fell below 1 just-noticeable difference (JND) and 100% fell below 2 JNDs]. This means that under lower light levels, model spectra would be an even better match to real wings. For the UV+ treatment, only ventral yellow was an excellent match to the *H. erato* ventral yellow for either *H. erato* sex. From this, we conclude that the Y+ paper model bears a strong resemblance to real *H. erato* yellow wings and the Y– paper model bears a strong resemblance to real *Eueides* yellow wings for *H. erato* butterflies under the experimental illuminant conditions in which they were tested.

For the UV-type and VS-type avian visual systems, the match between Y+ and UV+ and *H. erato* ventral yellow and between Y– and UV– *Eueides* ventral yellow was less good than if these same stimuli were viewed by the butterflies (Table 2). These results

indicate that for birds at least, under forest shade or edge illumination, no pair of stimuli fully captured the spectral differences between *Heliconius* or *Eueides* yellow wing colors. All pairs of model spectra used in behavioral experiments, however, differed by >1 JND for both birds and butterflies (except for Y+ versus Y– for ventral yellow viewed through the male eye; Table 3). This indicates that for both birds and butterflies, there was sufficient difference between the four model types to potentially elicit a behavioral response in the experiments described below.

Experiment 1: effect of model type on mate preference

To determine how *Heliconius* yellow and UV affect conspecific recognition, we presented wild-caught *H. erato* butterflies with artificial butterfly models that had manipulated yellow and UV coloration. Preference toward models was measured in the form of approaches and courtship events. We found a strong model type effect on the number of butterfly approaches toward 3-OHK yellow and UV models. There were significantly more approaches toward Y+ than Y– models (two-way ANOVA, $F=16.287$, $P<0.0001$, $n=80$), and toward UV+ than UV– models ($F=10.469$, $P=0.002$, $n=80$; Fig. 2A, black lines). There was no apparent effect of sex on butterfly approach behavior ($F=2.738$, $P=0.099$, $n=80$ for Y; $F=0.049$, $P=0.952$, $n=80$ for UV), suggesting that males and females approach the models at equal rates. Specific male and female behaviors for all comparisons are illustrated in Fig. S4.

Regarding courtship behavior, we found a strong model type effect where Y+ models were courted much more than Y– models ($F=11.731$, $P=0.0008$, $n=80$; Fig. 2A, red lines). The test for the main effect of sex shows that males court Y models at a significantly higher rate than females ($F=9.211$, $P=0.0002$, $n=80$). However, we found no significant model type effect on the number of courtship events directed toward UV+ and UV– models ($F=2.304$, $P=0.131$, $n=80$). There was also no effect of sex on butterfly courtship behavior toward the UV models ($F=0.701$, $P=0.498$, $n=80$).

Table 3. JNDs between model spectra through the eyes of male and female *H. erato* and representatives of the UV- and VS-type bird visual systems

	Y+ vs Y–				UV+ vs UV–			
	Butterfly		Bird		Butterfly		Bird	
	Female	Male	UV	VS	Female	Male	UV	VS
Dorsal yellow	1.04	1.77	N/A	N/A	2.38	1.28	N/A	N/A
Dorsal red	N/A	N/A	N/A	N/A	2.09	1.22	N/A	N/A
Ventral yellow	1.27	0.14	3.11 (3.37)	1.86 (1.91)	2.42	1.23	5.04 (5.38)	0.97 (1.03)
Ventral red	N/A	N/A	N/A	N/A	2.28	1.23	4.73 (5.05)	1.01

For butterflies, sunny cage illumination and for birds, partial forest cover illumination was used. Numbers in parentheses represent spectra modeled with forest edge illumination.

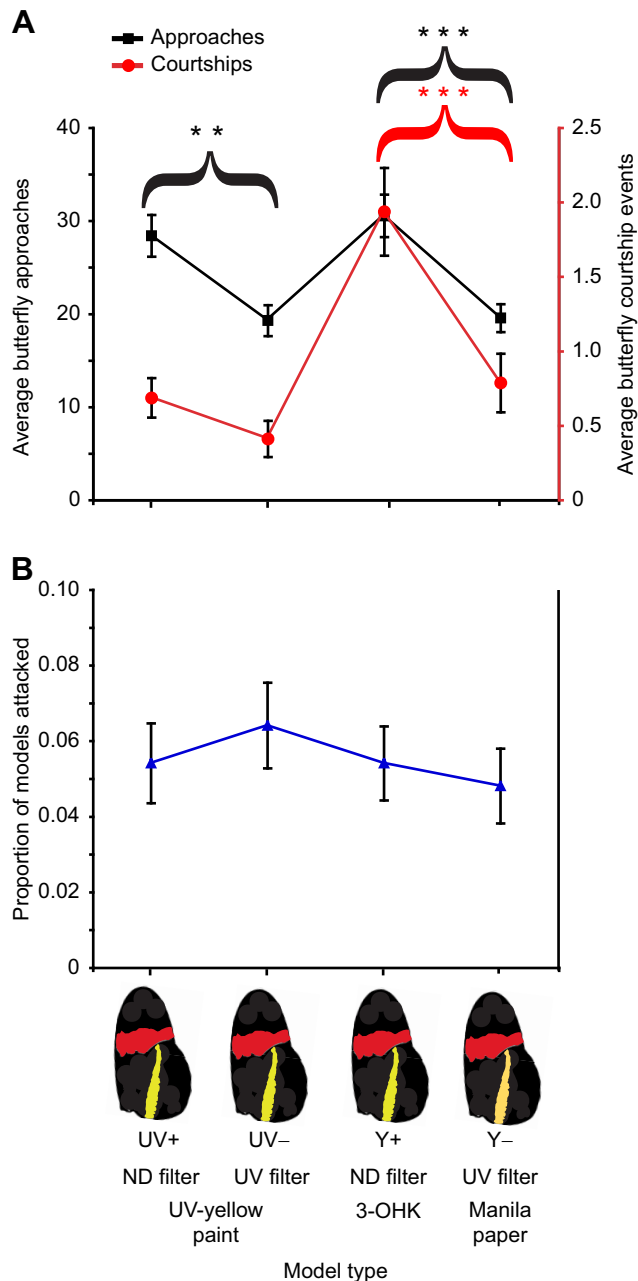


Fig. 2. UV- and 3-OHK-manipulated butterfly models experience different rates of approach and courtship behavior by butterflies and similar rates of predation by birds. The four model types differ according to whether UV-yellow paint (UV+, UV–), 3-OHK pigment (Y+) or Manila paper (Y–) was used to produce the yellow hindwing bar and according to whether a neutral-density (ND) filter (+treatments) or a UV-blocking filter (–treatments) was used. (A) Mean (\pm s.e.m.) approach (left axis, black) and courtship (right axis, red) values (each $n=80$ butterflies: 40 males and 40 females). Asterisks represent the P -values from pairwise comparisons (two-way ANOVA), where * $P<0.05$, ** $P<0.01$, *** $P<0.001$. (B) Mean (\pm s.e.m.) proportion of models attacked at each site (total $n=2000$: 500 of each model type, 100 sites). Pairwise comparisons (zero-inflated Poisson regression model with a two-tailed estimate) show all $P>0.05$.

Experiment 2: predator response to 3-OHK yellow and UV in different forest habitats

Previously, we showed that birds preferentially attack achromatic *H. erato* models over Y+ chromatic models (Fig. S1) (Finkbeiner et al., 2014), as expected if chromatic cues serve as aposematic

signals to avian predators. To test whether birds differentially attack yellow- or UV-manipulated models, predation was measured as the frequency of avian attacks on models in the forest. A total of 110 avian attacks were recorded (over 4 days of predator exposure for 500 models of each type): 27 and 24 attacks on Y+ and Y– models, and 27 and 32 attacks on UV+ and UV– models, respectively. Using a zero-inflated Poisson regression model, we detected no difference in predation between Y+ and Y– models: (z -value= -0.014 , $P=0.989$, $n=1000$; Fig. 2B), and no difference in predation between UV+ and UV– models: (z -value= -0.536 , $P=0.592$, $n=1000$; Fig. 2B). A test of whether forest type affected predator behavior found no difference in predation between the model types in forest cover, forest edge, Pipeline Road (unpaved road with partial forest cover) and paved road with partial forest cover (all $P>0.10$). Although our prior experiments indicated that avian predators differentially attack *H. erato* paper models that differ in both red and yellow color and pattern (Finkbeiner et al., 2014), the results presented here indicate that avian predators do not differentially attack 3-OHK yellow and other yellow or UV+ and UV– models in field trials.

Fluorescence does not contribute to the yellow signal

The absorption spectrum of 3-OHK has a distinctive peak (λ_{\max}) at 380 nm (Fig. 3C), so this wavelength was chosen as the excitation wavelength for fluorescence measurements (10 nm bandwidth). The excitation spectrum of the pigment (Fig. 3D, black line) is in full agreement with absorption measurements demonstrating that 380 nm is the peak excitation wavelength. The fluorescence of the pigment has a broad spectrum, with the peak of the emission around 508 nm (Fig. 3D, green line). Notably, the emission spectrum of 3-OHK overlaps well with the visible portion of *Heliconius* yellow, suggesting the fluorescence of 3-OHK might in principle contribute to the signal in the visible range.

In order to measure the efficiency of this emission, and hence understand whether the fluorescence might contribute significantly to the signal, we determined the fluorescence quantum yield of 3-OHK. Quantum yield is characterized as the ratio of the number of photons emitted to the number of photons absorbed (Williams et al., 1983; Nad and Pal, 2003). Quantum yield of 3-OHK was obtained by comparing 3-OHK with a standard and well-characterized fluorescent molecule, Coumarin 500 (Dhami et al., 1995), which has similar absorbance and fluorescence peaks to 3-OHK (Fig. S5). We were therefore surprised that the quantum yield of 3-OHK in methanol indicated that the emission is unlikely to be visible under normal illumination (quantum yield= 5.1×10^{-4}). By contrast, the quantum yield of our standard Coumarin 500 was 0.46 (Nad and Pal, 2003) or nearly a thousand times brighter than 3-OHK under similar conditions.

To be certain that these conclusions for 3-OHK in solution would also apply to 3-OHK on real wings in daylight illumination, additional experiments were carried out. Reflectance spectra of *H. erato* wings with and without a neutral-density filter (Mylar film) or a 400 nm cut-off filter (UV film), using a 150 W xenon arc lamp as a light source (which has a spectrum that resembles daylight illumination), were measured. If 3-OHK fluorescence does not contribute to the *Heliconius* yellow signal in broad-spectrum light, then measurements of *H. erato* wing reflectance spectra using a UV-cut off filter, which blocks excitation, should have no effect on the measured spectra in the visible range. That is indeed what we observed (Fig. 4). This series of experiments leads us to conclude that fluorescence does not contribute to the 3-OHK visual signal under broad-spectrum illumination.

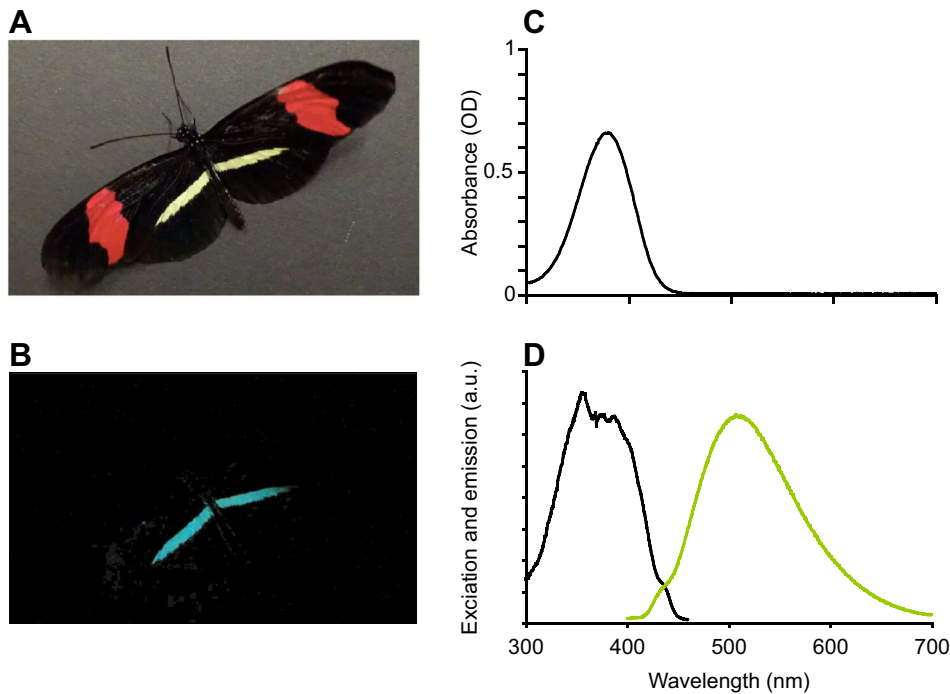


Fig. 3. *Heliconius erato* fluorescence and 3-hydroxy-DL-kynurenine (3-OHK) absorption, excitation, and emission spectra. (A,B) Adult *H. erato* photographed under (A) white light and (B) UV illumination to induce fluorescence. (C) Absorption spectrum of 3-OHK in methanol ($\lambda_{\text{max}}=380$ nm). The y-axis is in units of optical density (OD). (D) Excitation and emission spectrum of 3-OHK. The y-axis is in arbitrary units (a.u.). Emission has a broad spectrum with a peak around 508 nm. The absorption, excitation and emission spectra of three different 3-OHK dilutions in methanol were each measured once. Shown are spectra within the linear range of detection.

DISCUSSION

3-OHK coloration is preferred by *H. erato*

Butterflies are astonishingly diverse in their coloration, but the phylogenetic origins of new pigimentary coloration and the evolutionary forces that may have governed the adoption of a new pigment have rarely been investigated. Previously, we showed that 3-OHK pigmentation is a synapomorphy of the genus *Heliconius*, being an ancestral character for the genus, but absent for sister genera such as *Eueides* (Briscoe et al., 2010). Here, we have attempted to investigate how 3-OHK pigmentation functions as a signal for *H. erato* mate choice and defense. *Heliconius* yellow

coloration has a spectrum that includes reflectance maxima in the UV and human-visible range as well as fluorescence (Fig. 1A,B, Fig. 3A–D). Evidence here indicates that both the UV and long-wavelength components of the reflectance spectrum contribute to the visual signal *H. erato* butterflies use for conspecific recognition, but qualitatively that the UV part may be less important for *H. erato* courtship than it is for approach behavior. Specifically, the butterflies demonstrated clear preferences under all circumstances for Y+ over Y– (Fig. 2A). It is notable that our discriminability modeling of male and female *H. erato* vision indicates that for the butterflies at least the Y+ yellows are a good match to real *H. erato* yellow wing colors and Y– yellows are a good match to real *Eueides* yellow wing colors (Table 1). These results provide the first empirical evidence that *H. erato* butterflies prefer 3-OHK yellows to yellows found on the wings of their sister-genera, *Eueides*, and the first empirical evidence that the evolution of 3-OHK pigmentation in *Heliconius* may have been driven by sexual selection.

The interpretation of the UV+ and UV– treatments is a little less clear. The UV+ and UV– models had the same long-wavelength reflectance, but differed in the UV. UV+ models were approached by both sexes more frequently than UV– models, but while there was a trend towards preferring UV+ models during mating attempts, this difference was non-significant. This observation is perhaps surprising in view of the idea that at least for birds UV may be a short-range signal (Stevens and Cuthill, 2007). However, our discriminability calculations indicate that the UV+ dorsal yellow model color was not a good match to real *H. erato* dorsal yellow (Table 1). Neither the long-wavelength nor the UV reflectance for dorsal yellow UV treatments was as similar to natural *H. erato* dorsal yellow as was the Y+ treatment (Fig. 1A, Table 1). It may be that a closer match to the natural *H. erato* spectrum – including in the UV – is needed to elicit a stronger courtship response.

Many prior studies of butterfly mate choice have examined the preferences of one sex or the other but not both (Knüttel and Fiedler, 2001; Fordyce et al., 2002; Ellers and Boggs, 2003; Sweeney et al., 2003; Kemp, 2007b). We note that our mate preference results indicate equal responses to models by males and females with

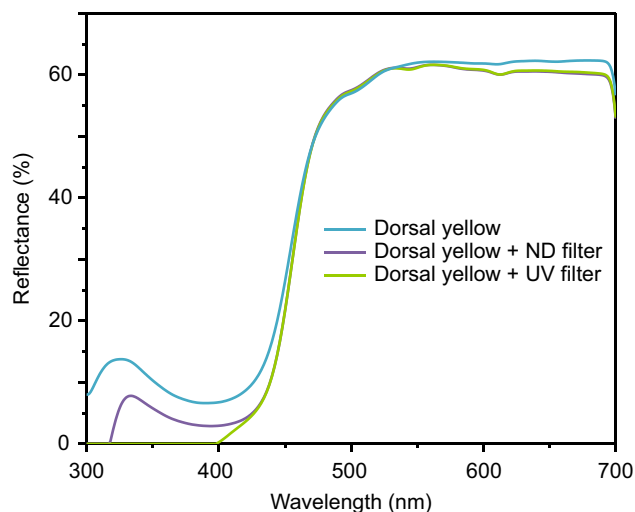


Fig. 4. Reflectance spectrum of a *H. erato* dorsal yellow hindwing bar with and without ND or UV-cutoff filters as measured using daylight-simulating illumination. The ND filter (Mylar) has an identical spectrum to the UV-cutoff filter in the visible range (above 400 nm), indicating that UV-induced fluorescence has no impact on the reflectance spectrum of 3-OHK yellow. Spectra from two different *H. erato* specimens were taken. Measurements from a single *H. erato* specimen are shown.

respect to approach behavior. This shows that females are ‘active’ during such preference studies (see Movies S2 and S3), and that females and males may share similar preferences for *Heliconius* yellow and UV in conspecifics. In nature, females may use approach behavior in non-mating-related interactions (Crane, 1955, 1957), such as following between pollen resources or to new roosting locations (Waller and Gilbert, 1982; Finkbeiner, 2014).

Our field study results show that 3-OHK yellow and UV do not alter avian predation rates in themselves, despite studies showing that birds use UV for mate recognition and foraging (Bennett et al., 1996; Siitari et al., 1999; Lyytinen et al., 2004). Recent work has shown that birds have even lower than expected UV sensitivity when looking at stimuli against a UV-poor background (Chavez et al., 2014) and understory-dwelling birds may have lower UV opsin expression than canopy-dwelling birds (Bloch, 2015). Our results resemble those of Lyytinen et al. (2000), who also found no support for UV as an aposematic signal for bird predators. Moreover, we provide experimental evidence that, in natural conditions, the mimicry between *Heliconius* yellow/UV coloration and non-*Heliconius* yellow/non-UV coloration in butterflies is successful for deterring birds. Given that we found no indication that *Heliconius* yellow and UV enhance aposematic signaling toward avian predators, this reinforces the notion that the phylogenetic switch from using other yellow pigments to 3-OHK as a signal on *Heliconius* wings is significant exclusively in relation to intraspecific communication.

Fluorescence does not function as a signal

Several studies have concluded that fluorescence is an important component of complex signals in aquatic animals because of the contrast between narrow-band down-welling blue light and long-wavelength fluorescence (Mazel et al., 2004; Gerlach et al., 2014). However, the evidence that fluorescence contributes to signaling in terrestrial animals, where the illumination spectrum is broadband, is much more limited and somewhat mixed. For instance, one laboratory study of fluorescence in budgerigars (*Melopsittacus undulatus*) suggested that fluorescence contributed to sexual signaling (Arnold et al., 2002) while two other studies of the same species did not (Pearn et al., 2001, 2003). In spiders, lab studies indicate that fluorescence plays a role in male mate choice while UV plays a role in female mate choice (Lim et al., 2007). A paper investigating UV and fluorescence in damselfly signaling (Guillermo-Ferreira et al., 2014) concluded that there might be a possible contribution of fluorescence to the signal; however, important controls necessary to confirm this were absent.

To our knowledge, we report here for the first time that the yellow wing coloration of *Heliconius* is fluorescent (Fig. 3), although Rawson (1968) mentions anecdotally that *H. erato* and *H. charithonia* wings are fluorescent but without specifying that it is the yellow portion of the wings, and without identifying the fluorescent chemical. However, by measuring the absorption, excitation and emission spectra and quantum yield of 3-OHK, together with wing reflectance spectra using daylight-simulating illumination, we found no evidence that 3-OHK fluorescence enhances the reflectance spectrum of *Heliconius* yellow under broad-band illumination. Although the spectral sensitivity of the *H. erato* blue receptor (470 nm) is well-suited to detecting 3-OHK fluorescence (McCulloch et al., 2016) we found no evidence that under natural illumination, fluorescence contributes to the 3-OHK signal in the visible range. Our result highlights the importance of quantifying fluorescence using several methods, and specifically under broad-band daylight-simulating illumination,

before concluding that it contributes to a signal under terrestrial environments (e.g. Andrews et al., 2007).

Conclusions

In summary, we demonstrate that *Heliconius* butterflies prefer 3-OHK yellow pigments in the context of conspecific signaling, these pigments have likely been selected for their reflectance properties in the visible range, and that fluorescence does not contribute to the visual signal. These results advance our understanding of the selective forces driving the transition from using other yellow pigments to using 3-OHK pigmentation in the genus *Heliconius*. We provide strong evidence that 3-OHK pigmentation is maintained because it allows *Heliconius* species to recognize conspecifics for interspecific communication and sexual selection, whilst retaining the potential benefits of Müllerian mimicry with genera such as *Eueides*.

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Competing interests

The authors declare no competing or financial interests.

Author contributions

S.D.F. designed butterfly models, carried out and analyzed field predation and mate preference experiments, and wrote the manuscript; D.A.F. contributed measurements and analysis of physical fluorescence properties; D.O. and A.D.B. conceived the study and edited the manuscript; A.D.B. designed butterfly models, calculated discriminabilities, performed experiments, analyzed fluorescence data and wrote the manuscript. All authors gave final approval for publication.

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Data Availability

Data are available from the Dryad Digital Repository (Finkbeiner et al., 2017): <http://dx.doi.org/10.5061/dryad.n0v5m>.

Supplementary information

Supplementary information available online at <http://jeb.biologists.org/lookup/doi/10.1242/jeb.153593.supplemental>

References

- Andrews, K., Reed, A. M. and Masta, S. E. (2007). Spiders fluoresce variably across many taxa. *Biol. Lett.* **3**, 265–267.
- Arikawa, K., Wakakuwa, M., Qiu, X., Kurasawa, M. and Stavenga, D. G. (2005). Sexual dimorphism of short-wavelength photoreceptors in the small white butterfly, *Pieris rapae crucivora*. *J. Neurosci.* **25**, 5935–5942.
- Arnold, K. E., Owens, I. P. F. and Marshall, N. J. (2002). Fluorescent signaling in parrots. *Science* **295**, 92.
- Bennett, A. T. D., Cuthill, I. C., Partridge, J. C. and Maier, E. J. (1996). Ultraviolet vision and mate choice in zebra finches. *Nature* **380**, 433–435.
- Benson, W. W. (1972). Natural selection for Müllerian mimicry in *Heliconius erato* in Costa Rica. *Science* **176**, 936–939.
- Bloch, N. I. (2015). Evolution of opsin expression in birds driven by sexual selection and habitat. *Proc. R. Soc. Lond. B.* **282**, 20142321.
- Briscoe, A. D. and Bernard, G. D. (2005). Eyeshine and spectral tuning of long wavelength-sensitive rhodopsins: No evidence for red-sensitive photoreceptors among five Nymphalini butterfly species. *J. Exp. Biol.* **208**, 687–696.
- Briscoe, A. D., Bybee, S. M., Bernard, G. D., Yuan, F., Sison-Mangus, M. P., Reed, R. D., Warren, A. D., Llorente-Bousquets, J. and Chiao, C.-C. (2010). Positive selection of a duplicated UV-sensitive visual pigment coincides with wing

- pigment evolution in *Heliconius* butterflies. *Proc. Natl. Acad. Sci. USA* **107**, 3628–3633.
- Bybee, S. M., Yuan, F., Ramstetter, M. D., Llorente-Bousquets, J., Reed, R. D., Osorio, D. and Briscoe, A. D. (2012). UV photoreceptors and UV-yellow wing pigments in *Heliconius* butterflies allow a color signal to serve both mimicry and intraspecific communication. *Am. Nat.* **179**, 38–51.
- Chai, P. (1986). Field observations and feeding experiments on the responses of rufous-tailed jacamars (*Galbula ruficauda*) to freeflying butterflies in a tropical rainforest. *Biol. J. Linn. Soc.* **29**, 161–189.
- Chavez, J., Kelber, A., Vorobyev, M. and Lind, O. (2014). Unexpectedly low UV-sensitivity in a bird, the budgerigar. *Biol. Lett.* **10**, 20140670.
- Chen, P.-J., Arikawa, K. and Yang, E.-C. (2013). Diversity of the photoreceptors and spectral opponency in the compound eye of the golden birdwing, *Troides aeacus formosanus*. *PLoS ONE* **8**, e62240.
- Crane, J. (1955). Imaginal behavior of a Trinidad butterfly, *Heliconius erato hydara* Hewitson, with special reference to the social use of color. *Zoologica* **40**, 167–196.
- Crane, J. (1957). Imaginal behavior in butterflies of the family Heliconiidae: changing social patterns and irrelevant actions. *Zoologica* **42**, 135–145.
- Cronin, T. W. and Bok, M. J. (2016). Photoreception and vision in the ultraviolet. *J. Exp. Biol.* **219**, 2790–2801.
- Cronin, T. W., Johnsen, S., Marshall, N. J. and Warrant, E. J. (2014). *Visual Ecology*, pp. 432. Princeton, NJ, USA: Princeton University Press.
- Cummings, M. E., Rosenthal, G. G. and Ryan, M. J. (2003). A private ultraviolet channel in visual communication. *Proc. R. Soc. Lond. B* **270**, 897–904.
- Dell’Aglia, D. D., Stevens, M. and Jiggins, C. D. (2016). Avoidance of an aposematically coloured butterfly by wild birds in a tropical forest. *Ecol. Entomol.* **41**, 627–632.
- Detto, T. and Blackwell, P. R. Y. (2009). The fiddler crab *Uca mjoebergi* uses ultraviolet cues in mate choice but not aggressive interactions. *Anim. Behav.* **78**, 407–411.
- Dhami, S., de Mello, A. J., Rumbles, G., Bishop, S. M., Phillips, D. and Beeby, A. (1995). Phthalocyanine fluorescence at high concentration: dimers or reabsorption effect? *Photochem. Photobiol.* **61**, 341.
- Eguchi, E. and Meyer-Rochow, V. B. (1983). Ultraviolet photography of forty-three species of Lepidoptera representing ten families. *Annot. Zool. Jap.* **56**, 10–18.
- Ellers, J. and Boggs, C. L. (2003). The evolution of wing color: male mate choice opposes adaptive wing color divergence in *Colias* butterflies. *Evolution* **57**, 1100–1106.
- Finkbeiner, S. D. (2014). Communal roosting in *Heliconius* butterflies (Nymphalidae): roost recruitment, establishment, fidelity, and resource use trends based on age and sex. *J. Lep. Soc.* **68**, 10–16.
- Finkbeiner, S. D., Fishman, D. A., Osorio, D. and Briscoe, A. D. (2017). Data from: Ultraviolet and yellow reflectance but not fluorescence is important for visual discrimination of conspecifics by *Heliconius erato*. *Dryad Digital Repository*. <http://dx.doi.org/10.5061/dryad.n0v5m>.
- Finkbeiner, S. D., Briscoe, A. D. and Reed, R. D. (2012). The benefit of being a social butterfly: communal roosting deters predation. *Proc. R. Soc. Lond. B* **279**, 2769–2776.
- Finkbeiner, S. D., Briscoe, A. D. and Reed, R. D. (2014). Warning signals are seductive: relative contributions of color and pattern to predator avoidance and mate attraction in *Heliconius* butterflies. *Evolution* **68**, 3410–3420.
- Fordyce, J. A., Nice, C. C., Forister, M. L. and Shapiro, A. M. (2002). The significance of wing pattern diversity in the Lycaenidae: mate discrimination by two recently diverged species. *J. Evol. Biol.* **15**, 871–879.
- Frentiu, F. D. and Briscoe, A. D. (2008). A butterfly’s eye view of birds. *BioEssays* **30**, 1151–1162.
- Gerlach, T., Sprenger, D. and Michiels, N. K. (2014). Fairy wrasses perceive and respond to their deep red fluorescent coloration. *Proc. R. Soc. Lond. B* **281**, pii: 20140787.
- Ghiradella, H. (1974). Development of ultraviolet-reflecting butterfly scales: how to make an interference filter. *J. Morphol.* **142**, 395–410.
- Grether, G. F., Kolluru, R. and Nersissian, K. (2004). Individual colour patches as multicomponent signals. *Biol. Rev.* **79**, 583–610.
- Guillermo-Ferreira, R., Therézio, E. M., Gehlen, M. H., Bispo, P. C. and Marletta, A. (2014). The role of wing pigmentation, UV and fluorescence as signals in a neotropical damselfly. *J. Insect Behav.* **27**, 67–80.
- Hart, N. S., Partridge, J. C., Cuthill, I. C. and Bennett, A. T. D. (2000). Visual pigments, oil droplets, ocular media and cone photoreceptor distribution in two species of passerine bird: the blue tit (*Parus caeruleus* L.) and the blackbird (*Turdus merula* L.). *J. Comp. Physiol. A* **186**, 375–387.
- Jackman, S. (2011). *pscl: Classes and Methods for R Developed in the Political Science Computational Laboratory*, Stanford University. Stanford, California: Department of Political Science, Stanford University. R package version 1.04.1.
- Johnsen, A., Andersson, S., Örnborg, J. and Lifjeld, J. T. (1998). Ultraviolet plumage ornamentation affects social mate choice and sperm competition in blue throats (Aves: *Luscinia s. svecica*): a field experiment. *Proc. R. Soc. Lond. B* **265**, 1313–1318.
- Kelber, A., Vorobyev, M. and Osorio, D. (2003). Animal colour vision: behavioural tests and physiological concepts. *Biol. Rev.* **78**, 81–118.
- Kemp, D. J. (2007a). Female mating biases for bright ultraviolet iridescence in the butterfly *Eurema hecabe* (Pieridae). *Behav. Ecol.* **19**, 1–8.
- Kemp, D. J. (2007b). Female butterflies prefer males bearing bright iridescent ornamentation. *Proc. R. Soc. Lond. B* **274**, 1043–1047.
- Kemp, D. J. (2008). Female mating biases for bright ultraviolet iridescence in the butterfly *Eurema hecabe* (Pieridae). *Behav. Ecol.* **19**, 1–8.
- Knüttel, H. and Fiedler, K. (2001). Host-plant-derived variation in ultraviolet wing patterns influences mate choice by male butterflies. *J. Exp. Biol.* **204**, 2447–2459.
- Koshitaka, H., Kinoshita, M., Vorobyev, M. and Arikawa, K. (2008). Tetrachromacy in a butterfly that has eight varieties of spectral receptors. *Proc. R. Soc. Lond. B* **275**, 947–954.
- Lagorio, M. G., Cordon, G. B. and Iriel, A. (2015). Reviewing the relevance of fluorescence in biological systems. *Photochem. Photobiol. Sci.* **14**, 1538–1559.
- Lim, L. M. L., Land, M. F. and Li, D. (2007). Sex-specific UV and fluorescence signals in jumping spiders. *Science* **315**, 481.
- Lind, O. and Kelber, A. (2009). Avian colour vision: effects of variation in receptor sensitivity and noise data on model predictions as compared to behavioural results. *Vision Res.* **49**, 1939–1947.
- Llaurens, V., Joron, M. and Théry, M. (2014). Cryptic differences in colour among Müllerian mimics: how can the visual capacities of predators and prey shape the evolution of wing colours? *J. Evol. Biol.* **27**, 531–540.
- Lyytinen, A., Alatalo, R. V., Lindström, L. and Mappes, J. (2000). Can ultraviolet cues function as aposematic signals? *Behav. Evol.* **12**, 65–70.
- Lyytinen, A., Lindström, L. and Mappes, J. (2004). Ultraviolet reflection and predation risk in diurnal and nocturnal Lepidoptera. *Behav. Evol.* **15**, 982–987.
- Mazel, C. H., Cronin, T. W., Caldwell, R. L. and Marshall, N. J. (2004). Fluorescent enhancement of signaling in a mantis shrimp. *Science* **303**, 51.
- McCulloch, K. J., Osorio, D. and Briscoe, A. D. (2016). Sexual dimorphism in the compound eye of *Heliconius erato*: a nymphalid butterfly with at least five spectral classes of photoreceptor. *J. Exp. Biol.* **219**, 2377–2387.
- Meyer-Rochow, V. B. (1991). Differences in ultraviolet light patterns in the New Zealand lycaenid butterflies *Lycaena salustius*, *L. rauparaha*, and *L. feredayi* as a likely isolating mechanism. *J. Roy. Soc. New Zealand* **21**, 169–177.
- Nad, S. and Pal, H. (2003). Photophysical properties of Coumarin-500 (C500): unusual behavior in nonpolar solvents. *J. Phys. Chem. A* **107**, 501–507.
- Obara, Y., Koshitaka, H. and Arikawa, K. (2008). Better mate in the shade: enhancement of male mating behaviour in the cabbage butterfly, *Pieris rapae crucivora*, in a UV-rich environment. *J. Exp. Biol.* **211**, 3698–3702.
- Olsson, P., Lind, O. and Kelber, A. (2015). Bird colour vision: behavioral thresholds reveal receptor noise. *J. Exp. Biol.* **218**, 184–193.
- Painting, C. J., Rajamohan, G., Chen, Z., Zeng, H. and Li, D. (2016). It takes two peaks to tango: the importance of UVB and UVA in sexual signalling in jumping spiders. *Anim. Behav.* **113**, 137–146.
- Pearn, S. M., Bennett, A. T. D. and Cuthill, I. C. (2001). Ultraviolet vision, fluorescence and mate choice in a parrot, the budgerigar *Melopsittacus undulatus*. *Proc. R. Soc. Lond. B* **268**, 2273–2279.
- Pearn, S. M., Bennett, A. T. D. and Cuthill, I. C. (2003). The role of ultraviolet reflectance and ultraviolet-A induced fluorescence in the appearance of budgerigar plumage: insights from spectrofluorometry and reflectance spectrophotometry. *Proc. R. Soc. Lond. B* **270**, 859–865.
- Prum, R. O. and Torres, R. H. (2003). Structural colouration of avian skin: convergent evolution of coherently scattering dermal collagen arrays. *J. Exp. Biol.* **206**, 2409–2429.
- R Development Core Team (2010). *R: A language and environment for statistical computing*. Vienna, Austria: R Foundation for Statistical Computing.
- Rawson, G. W. (1968). Study of fluorescent pigments in Lepidoptera by means of paper partition chromatography. *J. Lep. Soc.* **22**, 27–40.
- Reed, R. D., McMillan, W. O. and Nagy, L. M. (2008). Gene expression underlying adaptive variation in *Heliconius* wing patterns: non-modular regulation of overlapping *cinnabar* and *vermillion* prepatterns. *Proc. R. Soc. Lond. B* **275**, 37–45.
- Robertson, K. A. and Monteiro, A. (2005). Female *Bicyclus anynana* butterflies choose males on the basis of their dorsal UV-reflective eyespot pupils. *Proc. R. Soc. Lond. B* **272**, 1541–1546.
- Rutowski, R. L. (1977). The use of visual cues in sexual and species discrimination by males of the small sulphur butterfly *Eurema lisa* (Lepidoptera, Pieridae). *J. Comp. Physiol.* **115**, 61–74.
- Rutowski, R. L., Macedonia, J. M., Morehouse, N. and Taylor-Taft, L. (2005). Pterin pigments amplify iridescent ultraviolet signal in males of the orange sulphur butterfly, *Colias eurytheme*. *Proc. R. Soc. Lond. B* **272**, 2329–2335.
- Shawkey, M. D. and Hill, G. E. (2005). Carotenoids need structural colours to shine. *Biol. Lett.* **1**, 121–124.
- Siitari, H., Honkava, J. and Viitala, J. (1999). Ultraviolet reflection of berries attracts foraging birds. A laboratory study with redwings (*Turdus iliacus*) and bilberries (*Vaccinium myrtillus*). *Proc. R. Soc. Lond. B* **266**, 2125–2129.
- Silberglied, R. E. and Taylor, O. R. (1978). Ultraviolet reflection and its behavioral role in the courtship of the sulfur butterflies *Colias eurytheme* and *C. philodice* (Lepidoptera, Pieridae). *Behav. Ecol. Sociobiol.* **3**, 203–243.
- Sison-Mangus, M. P., Briscoe, A. D., Zaccardi, G., Knüttel, H. and Kelber, A. (2008). The lycaenid butterfly *Polyommatus icarus* uses a duplicated blue opsin to see green. *J. Exp. Biol.* **211**, 361–369.

- Smith, E. J., Partridge, J. C., Parsons, K. N., White, E. M., Cuthill, I. C., Bennett, A. T. D. and Church, S. C. (2002). Ultraviolet vision and mate choice in the guppy (*Poecilia reticulata*). *Behav. Ecol.* **13**, 11–19.
- Stalleicken, J., Labhart, T. and Mouritsen, H. (2006). Physiological characterization of the compound eye in monarch butterflies with focus on the dorsal rim area. *J. Comp. Physiol. A* **192**, 321–331.
- Stavenga, D. G., Stowe, S., Siebke, K., Zeil, J. and Arikawa, K. (2004). Butterfly wing colours: scale beads make white pierid wings brighter. *Proc. R. Soc. Lond. B* **271**, 1577–1584.
- Stavenga, D. G., Leertouwer, H. L., Marshall, N. J. and Osorio, D. (2011). Dramatic colour changes in a bird of paradise caused by uniquely structured breast feather barbules. *Proc. R. Soc. Lond. B* **278**, 2098–2104.
- Stavenga, D. G., Leertouwer, H. L. and Wilts, B. D. (2014). Coloration principles of nymphaline butterflies - thin films, melanin, ommochromes and wing scale stacking. *J. Exp. Biol.* **217**, 2171–2180.
- Stevens, M. and Cuthill, I. C. (2007). Hidden messages: Are ultraviolet signals a special channel in avian communication? *Bioscience* **57**, 501–507.
- Sweeney, A., Jiggins, C. and Johnsen, S. (2003). Polarized light as a butterfly mating signal. *Nature* **423**, 31–32.
- Tokuyama, T., Senoh, S., Sakan, T., Brown, K. S. and Witkop, B. (1967). The photoreduction of kynurenic acid to kynurenine yellow and the occurrence of 3-hydroxy-L-kynurenine in butterflies. *J. Am. Chem. Soc.* **89**, 1017–1021.
- Toomey, M. B., Lind, O., Frederiksen, R., Curley, R. W., Riedl, K. M., Wilby, D., Schwartz, S. J., Witt, C. C., Harrison, E. A., Roberts, N. W. et al. (2016). Complementary shifts in photoreceptor spectral tuning unlock the full adaptive potential of ultraviolet vision in birds. *eLife* **5**, e15675.
- Vorobyev, M. and Osorio, D. (1998). Receptor noise as a determinant of colour thresholds. *Proc. R. Soc. Lond. B* **265**, 351–358.
- Vukusic, P. and Hooper, I. (2005). Directionally controlled fluorescence emission in butterflies. *Science* **310**, 1151.
- Vukusic, P. and Sambles, J. R. (2003). Photonic structures in biology. *Nature* **424**, 853–855.
- Waller, D. A. and Gilbert, L. E. (1982). Roost recruitment and resource utilization: observations on *Heliconius charithonia*. *J. Lep. Soc.* **36**, 178–184.
- Watt, W. B. (1964). Pteridine components of wing pigmentation in the butterfly *Colias eurytheme*. *Nature* **201**, 1326–1327.
- Williams, A. T. R., Winfield, S. A. and Miller, J. N. (1983). Relative fluorescence quantum yields using a computer controlled luminescence spectrometer. *Analyst* **108**, 1067.
- Yuan, F., Bernard, G. D., Le, J. and Briscoe, A. D. (2010). Contrasting modes of evolution of the visual pigments in *Heliconius* butterflies. *Mol. Biol. Evol.* **27**, 2392–2405.
- Zeileis, A., Kleiber, C. and Jackman, S. (2008). Regression models for count data in R. *J. Stat. Softw.* **27**, 1–25.